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Patents Act 1990

PROVISIONAL SPECIFICATION

Applicants:

THE UNIVERSITY OF QUEENSLAND

Invention Title:

TREATMENT OF OSTEOARTHRITIS

The invention is described in the following statement:

TREATMENT OF OSTEOARTHRITIS

FIELD OF THE INVENTION

This invention relates to the treatment of osteoarthritis with novel cyclic peptidic and peptidomimetic compounds which have the ability to modulate the activity of G protein-coupled receptors. The compounds preferably act as antagonists of the C5a receptor, and are active against C5a receptors on polymorphonuclear leukocytes and macrophages.

BACKGROUND OF THE INVENTION

All references, including any patents or patent applications, cited in this specification are hereby incorporated by reference. No admission is made that any reference constitutes prior art. The discussion of the references states what their authors assert, and the applicants reserve the right to challenge the accuracy and pertinency of the cited documents. It will be clearly understood that, although a number of prior art publications are referred to herein, this reference does not constitute an admission that any of these documents forms part of the common general knowledge in the art, in Australia or in any other country.

G protein-coupled receptors are prevalent throughout the human body, comprising approximately 60% of known cellular receptor types, and mediate signal transduction across the cell membrane for a very wide range of endogenous ligands. They participate in a diverse array of physiological and pathophysiological processes, including, but not limited to, those associated with cardiovascular, central and peripheral nervous system, reproductive, metabolic, digestive, immunological, inflammatory, and growth disorders, as well as other cell-regulatory and proliferative disorders. Agents which selectively modulate functions of G protein-coupled receptors have important therapeutic applications. These

receptors are becoming increasingly recognised as important drug targets, due to their crucial roles in signal transduction (G protein-coupled Receptors, IBC Biomedical Library Series, 1996).

One of the most intensively studied G protein-coupled receptors is the receptor for C5a. C5a is one of the most potent chemotactic agents known, and recruits neutrophils and macrophages to sites of injury, alters their morphology; induces degranulation; increases calcium mobilisation, vascular permeability (oedema) and neutrophil adhesiveness; contracts smooth muscle; stimulates release of inflammatory mediators, including histamine, TNF-α, IL-1, IL-6, IL-8, prostaglandins, and leukotrienes, and of lysosomal enzymes; promotes formation of oxygen radicals; and enhances antibody production (Gerard and Gerard, 1994).

Agents which limit the pro-inflammatory actions of C5a have potential for inhibiting chronic inflammation, and its accompanying pain and tissue damage. For these reasons, molecules which prevent C5a from binding to its receptors are useful for treating chronic inflammatory disorders driven by complement activation.

In our previous application No.PCT/AU98/00490, we described the three-dimensional structure of some analogues of the C-terminus of human C5a, and used this information to design novel compounds which bind to the human C5a receptor (C5aR), behaving as either agonists or antagonists of C5a. It had previously been thought that a putative antagonist might require both a C-terminal arginine and a C-terminal carboxylate for receptor binding and antagonist activity (Konteatis *et al*, 1994). In PCT/AU98/00490 we showed that in fact a terminal carboxylate group is not generally required either for high affinity binding to C5aR or for antagonist activity. Instead we found that a hitherto unrecognised structural feature, a turn conformation, was the key recognition feature for high affinity binding to the human C5a

receptor on neutrophils. As described in our Australian provisional application No. PR8334, filed on 17th October 2001, we used these findings to design constrained structural templates which enable hydrophobic groups to be assembled into a hydrophobic array for interaction with a C5a receptor. The entire disclosures of these specifications are incorporated herein by this reference.

Osteoarthritis is a non-inflammatory, chronic degenerative joint condition, characterized by degeneration of articular cartilage; in advanced cases there is also hypertrophy of bone at the joint margins, and changes in the synovial membrane. Secondary changes in underlying bone cause pain and affect joint function. At present, therapies available to treat osteoarthritis are limited to the use of analgesics or anti-inflammatory agents, reduction in pressure across the joint, and weight loss. Most current treatments are designed only to relieve pain and/or inflammation, and to reduce or prevent the disability caused by bone and cartilage degeneration. COX-II inhibitors such as Celebrex, Vioxx and Bextra, which target inflammation, have recently become available for the treatment of this condition.

The available drug therapies target the symptoms but not the underlying cause of this disease; none of them inhibits the degenerative structural changes which are responsible for its progression. The disease continues to progress, and total joint replacement, especially of the hip or knee, is ultimately necessary in many patients. Furthermore, clinical testing of new therapies is complicated by the fact that the disease manifests itself differently in each person.

A variety of agents, ranging from tumour necrosis factor antagonists to dietary supplements such as S-adenosyl methionine or boron compounds, are in various stages of clinical trial. However, there is a great need in the art for effective, non-toxic agents which do not require administration by injection, and which can be

produced at reasonable cost. To our knowledge none of these approved or experimental agents, and in particular no small molecule agent, targets the C5a receptor.

Osteoarthritis is strongly age-related, with over 50% of people over the age of 70 being treated for this condition. It is also associated with obesity and with over-use injuries, and is common in former athletes who engaged in weight-bearing sports. It is currently estimated that in the United States 35 million people – 13 percent of the population – are 65 and older, and that more than half of these people have radiological evidence of osteoarthritis in at least one joint. By 2030, 20 percent of Americans – about 70 million people – will have passed their 65th birthday and will be at risk for osteoarthritis. It is estimated that there are about 103 million osteoarthritis sufferers in the European Union.

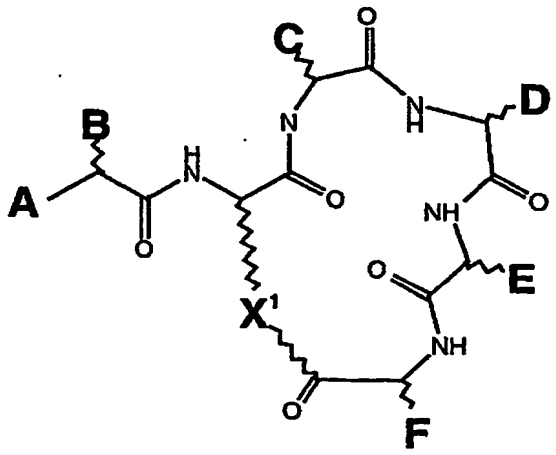
We now show for the first time that a specific inhibitor of the C5a receptor is able to ameliorate signs of damage in a model of induced osteoarthritis in dogs. This is the first reported case of an inhibitor of the complement system being used to modulate pathology in a model of osteoarthritis.

SUMMARY OF THE INVENTION

According to a first aspect, the invention provides a method of treatment of osteoarthritis, comprising the step of administering an effective amount of an inhibitor of a G protein-coupled receptor to a subject in need of such treatment.

Preferably the inhibitor is a compound which

(a) is an antagonist of a G protein-coupled receptor,
(b) has substantially no agonist activity, and
(c) is a cyclic peptide or peptidomimetic compound of formula I



where A is H, alkyl, aryl, NH_2, NH-alkyl, $N(alkyl)_2$, NH-aryl, NH-acyl, NH-benzoyl, $NHSO_3$, $NHSO_2$-alkyl, $NHSO_2$-aryl, OH, O-alkyl, or O-aryl;

B is an alkyl, aryl, phenyl, benzyl, naphthyl or indole group, or the side chain of a D- or L-amino acid such as L-phenylalanine or L-phenylglycine, but is not the side chain of glycine, D-phenylalanine, L-homophenylalanine, L-tryptophan, L-homotryptophan, L-tyrosine, or L-homotyrosine;

C is a small substituent, such as the side chain of a D-, L- or homo-amino acid such as glycine, alanine, leucine, valine, proline, hydroxyproline, or thioproline, but is preferably not a bulky substituent such as isoleucine, phenylalanine, or cyclohexylalanine;

D is the side chain of a neutral D-amino acid such as D-Leucine, D-homoleucine, D-cyclohexylalanine, D-homocyclohexylalanine, D-valine, D-norleucine, D-homo-norleucine, D-phenylalanine, D-tetrahydroisoquinoline, D-glutamine, D-glutamate, or D-tyrosine, but is preferably not a small substituent such as the side chain of glycine or D-alanine, a bulky planar side chain such as D-tryptophan, or a bulky charged side chain such as D-arginine or D-Lysine;

E is a bulky substituent, such as the side chain of an amino acid selected from the group consisting of L-

phenylalanine, L-tryptophan and L-homotryptophan, or is L-1-napthyl or L-3-benzothienyl alanine, but is not the side chain of D-tryptophan, L-N-methyltryptophan, L-homophenylalanine, L-2-naphthyl L-tetrahydroisoquinoline, L-cyclohexylalanine, D-leucine, L-fluorenylalanine, or L-histidine;

F is the side chain of L-arginine, L-homoarginine, L-citrulline, or L-canavanine, or a bioisostere thereof, ie. a side chain in which the terminal guanidine or urea group is retained, but the carbon backbone is replaced by a group which has different structure but is such that the side chain as a whole reacts with the target protein in the same way as the parent group; and

X is $-(CH_2)_nNH-$ or $(CH_2)_n-S-$, where n is an integer of from 1 to 4, preferably 2 or 3; $-(CH_2)_2O-$; $-(CH_2)_3O-$; $-(CH_2)_3-$; $-(CH_2)_4-$; $-CH_2COCHRNH-$; or $-CH_2-CHCOCHRNH-$, where R is the side chain of any common or uncommon amino acid.

In C, both the *cis* and *trans* forms of hydroxyproline and thioproline may be used.

Preferably A is an acetamide group, an aminomethyl group, or a substituted or unsubstituted sulphonamide group.

Preferably where A is a substituted sulphonamide, the substituent is an alkyl chain of 1 to 6, preferably 1 to 4 carbon atoms, or a phenyl or toluyl group.

In a particularly preferred embodiment, the compound has antagonist activity against C5aR, and has no C5a agonist activity.

The compound is preferably an antagonist of C5a receptors on human and mammalian cells including, but not limited to, human polymorphonuclear leukocytes and human macrophages. The compound preferably binds potently and selectively to C5a receptors, and more preferably has potent antagonist activity at sub-micromolar concentrations. Even more preferably the compound has a

receptor affinity IC50<25μM, and an antagonist potency IC50<1μM.

Most preferably the compound is selected from the group consisting of compounds **1** to **6**, **10** to **15**, **17**, **19**, **20**, **22**, **25**, **26**, **28**, **30**, **31**, **33** to **37**, **39** to **45**, **47** to **50**, **52** to **58** and **60** to **70** described in provisional application No.PR8334. In a particularly preferred embodiment, the compound is PMX53 (compound **1**), compound **33**, compound **60** or compound **45** described therein.

The inhibitor may be used in conjunction with one or more other agents for the treatment of osteoarthritis, including but not limited to analgesics such as aspirin, corticosteroids such as prednisolone, anti-inflammatory agents, including but not limited to non-steroidal anti-inflammatory agents such as diclofenac, naproxen or ketoprofen, or COX II inhibitors such as Celebrex, Vioxx, Bextra or eterocoxib, meloxicam, carprofen and the like. Other products used to treat osteoarthritis include glycosaminoglycans,pentosan polysulphate, eicosapentanoic acids, omega-3 fatty acids, chondroitin sulphate and glucosamine and intraarticular injections of hyaluronic acid.

The compositions of the invention may be formulated for oral, parenteral, inhalational, intranasal, rectal or transdermal use, but oral, injectable or percutaneous formulations are preferred. It is expected that most if not all compounds of the invention will be stable in the presence of metabolic enzymes, such as those of the gut, blood, lung or intracellular enzymes. Such stability can readily be tested by routine methods known to those skilled in the art.

Suitable formulations for administration by any desired route may be prepared by standard methods, for example by reference to well-known textbooks such as Remington: The Science and Practice of Pharmacy, Vol. II, 2000 (20th edition), A.R. Gennaro (ed), Williams & Wilkins, Pennsylvania.

While the invention is not in any way restricted to the treatment of any particular animal or species, it is particularly contemplated that the method of the invention will be useful in medical treatment of humans, and will also be useful in veterinary treatment, particularly of companion animals such as cats, dogs and birds, livestock such as cattle, horses, poultry and sheep, and zoo animals, including non-human primates, large bovids, felids, ungulates and canids.

The compound may be administered at any suitable dose and by any suitable route. Oral, parenteral or transdermal administration is preferred, because of the greater convenience and acceptability of these routes. The effective dose will depend on the nature of the condition to be treated, and the age, weight, and underlying state of health of the individual treatment. This will be at the discretion of the attending physician or veterinarian. Suitable dosage levels may readily be determined by trial and error experimentation, using methods which are well known in the art.

BRIEF DESCRIPTION OF THE FIGURES

Figure 1 shows the inhibition of the vascular leakage associated with a dermal Arthus reaction by intravenous (A), oral (B) and topical (C) AcF-[OPdChaWR], and appropriate controls (D).

Figure 2 shows the inhibition of the rise in circulating TNFα associated with a dermal Arthus reaction by intravenous (A), oral (B) and topical (C) AcF-[OPdChaWR], and appropriate topical controls (D).

Figure 3 shows the reduction of the pathology index associated with a dermal Arthus reaction by intravenous, oral and topical AcF-[OPdChaWR].

Figure 4 shows the inhibition of arthritic right knee joint swelling by AcF-[OPdChaWR] given orally on Days-2 to +14.

Figure 5 shows the inhibition of right knee joint TNFα and IL-6 levels in joint lavage. "Untreated" refers to an animal not treated with AcF-[OPdChaWR], but with its right knee challenged with antigen following sensitisation.

Figure 6a summarises the gait scores of dogs (n = 2) treated with placebo after transection of the cruciate ligament.

Figure 6b summarises the gait score of dogs (n = 2) treated with PMX53 (0.3mg/kg subcutaneously once a day) after transection of the cruciate ligament.

Figure 7 compares the histological appearance of knee joints from control and PMX-53 (3 D53)-treated dogs.

DETAILED DESCRIPTION OF THE INVENTION

For the purposes of this specification it will be clearly understood that the word "comprising" means "including but not limited to", and that the word "comprises" has a corresponding meaning.

As used herein, the singular forms "a", "an", and "the" include plural reference unless the context clearly dictates otherwise. Thus, for example, a reference to "an enzyme" includes a plurality of such enzymes, and a reference to "an amino acid" is a reference to one or more amino acids. Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any materials and methods similar or equivalent to those described herein can be used to practice or test the present invention, the preferred materials and methods are now described.

Abbreviations used herein are as follows:

D-Cha	D-cyclohexylamine
LPS	lipopolysaccharide
PMN	polymorphonuclear granulocyte
rp-HPLC	reverse phase-high performance liquid

	chromatography
TFA	trifluoroacetic acid;
Cit	citrulline
dCha	D-cyclohexylamine
DPhe	D-phenylalanine
ip	intraperitoneal
iv	intravenous
LPS	lipopolysaccharide
PMN	polymorphonuclear granulocyte
PMSF	phenylmethylsulfonyl fluoride
sc	subcutaneous

Throughout the specification conventional single-letter and three-letter codes are used to represent amino acids.

For the purposes of this specification, the term "alkyl" is to be taken to mean a straight, branched, or cyclic, substituted or unsubstituted alkyl chain of 1 to 6, preferably 1 to 4 carbons. Most preferably the alkyl group is a methyl group. The term "acyl" is to be taken to mean a substituted or unsubstituted acyl of 1 to 6, preferably 1 to 4 carbon atoms. Most preferably the acyl group is acetyl. The term "aryl" is to be understood to mean a substituted or unsubstituted homocyclic or heterocyclic aryl group, in which the ring preferably has 5 or 6 members.

A "common" amino acid is a L-amino acid selected from the group consisting of glycine, leucine, isoleucine, valine, alanine, phenylalanine, tyrosine, tryptophan, aspartate, asparagine, glutamate, glutamine, cysteine, methionine, arginine, lysine, proline, serine, threonine and histidine.

An "uncommon" amino acid includes, but is not restricted to, D-amino acids, homo-amino acids, N-alkyl amino acids, dehydroamino acids, aromatic amino acids other than phenylalanine, tyrosine and tryptophan, ortho-, meta- or para-aminobenzoic acid, ornithine, citrulline,

canavanine, norleucine, γ-glutamic acid, aminobutyric acid, L-fluorenylalanine, L-3-benzothienylalanine, and α,α-disubstituted amino acids.

Generally, the terms "treating", "treatment" and the like are used herein to mean affecting a subject, tissue or cell to obtain a desired pharmacological and/or physiological effect. The effect may be prophylactic in terms of completely or partially preventing a disease or sign or symptom thereof, and/or may be therapeutic in terms of a partial or complete cure of a disease.

"Treating" as used herein covers any treatment of, or prevention of disease in a vertebrate, a mammal, particularly a human, and includes: preventing the disease from occurring in a subject who may be predisposed to the disease, but has not yet been diagnosed as having it; inhibiting the disease, ie., arresting its development; or relieving or ameliorating the effects of the disease, ie., cause regression of the effects of the disease.

The invention includes the use of various pharmaceutical compositions useful for ameliorating disease. The pharmaceutical compositions according to one embodiment of the invention are prepared by bringing a compound of formula I, analogue, derivatives or salts thereof and one or more pharmaceutically-active agents or combinations of compound of formula I and one or more pharmaceutically-active agents into a form suitable for administration to a subject using carriers, excipients and additives or auxiliaries.

Frequently used carriers or auxiliaries include magnesium carbonate, titanium dioxide, lactose, mannitol and other sugars, talc, milk protein, gelatin, starch, vitamins, cellulose and its derivatives, animal and vegetable oils, polyethylene glycols and solvents, such as sterile water, alcohols, glycerol and polyhydric alcohols. Intravenous vehicles include fluid and nutrient replenishers. Preservatives include antimicrobial, anti-oxidants, chelating agents and inert gases. Other

pharmaceutically acceptable carriers include aqueous solutions, non-toxic excipients, including salts, preservatives, buffers and the like, as described, for instance, in Remington's Pharmaceutical Sciences, 20th ed. Williams & Wilkins (2000) and The British National Formulary 43rd ed. (British Medical Association and Royal Pharmaceutical Society of Great Britain, 2002; http://bnf.rhn.net), the contents of which are hereby incorporated by reference. The pH and exact concentration of the various components of the pharmaceutical composition are adjusted according to routine skills in the art. See Goodman and Gilman's The Pharmacological Basis for Therapeutics (7th ed., 1985).

The pharmaceutical compositions are preferably prepared and administered in dosage units. Solid dosage units include tablets, capsules and suppositories. For treatment of a subject, depending on activity of the compound, manner of administration, nature and severity of the disorder, age and body weight of the subject, different daily doses can be used. Under certain circumstances, however, higher or lower daily doses may be appropriate. The administration of the daily dose can be carried out both by single administration in the form of an individual dose unit or else several smaller dose units and also by multiple administration of subdivided doses at specific intervals.

The pharmaceutical compositions according to the invention may be administered locally or systemically in a therapeutically effective dose. Amounts effective for this use will, of course, depend on the severity of the disease and the weight and general state of the subject. Typically, dosages used *in vitro* may provide useful guidance in the amounts useful for *in situ* administration of the pharmaceutical composition, and animal models may be used to determine effective dosages for treatment of the cytotoxic side effects. Various considerations are described, eg. in Langer, Science, 249: 1527, (1990).

Formulations for oral use may be in the form of hard gelatin capsules, in which the active ingredient is mixed with an inert solid diluent, for example, calcium carbonate, calcium phosphate or kaolin. They may also be in the form of soft gelatin capsules, in which the active ingredient is mixed with water or an oil medium, such as peanut oil, liquid paraffin or olive oil.

Aqueous suspensions normally contain the active materials in admixture with excipients suitable for the manufacture of aqueous suspensions. Such excipients may be suspending agents such as sodium carboxymethyl cellulose, methyl cellulose, hydroxypropylmethylcellulose, sodium alginate, polyvinylpyrrolidone, gum tragacanth and gum acacia; dispersing or wetting agents, which may be (a) a naturally occurring phosphatide such as lecithin; (b) a condensation product of an alkylene oxide with a fatty acid, for example, polyoxyethylene stearate; (c) a condensation product of ethylene oxide with a long chain aliphatic alcohol, for example, heptadecaethylenoxycetanol; (d) a condensation product of ethylene oxide with a partial ester derived from a fatty acid and hexitol such as polyoxyethylene sorbitol monooleate, or (e) a condensation product of ethylene oxide with a partial ester derived from fatty acids and hexitol anhydrides, for example polyoxyethylene sorbitan monooleate.

The pharmaceutical compositions may be in the form of a sterile injectable aqueous or oleaginous suspension. This suspension may be formulated according to known methods using suitable dispersing or wetting agents and suspending agents such as those mentioned above. The sterile injectable preparation may also be a sterile injectable solution or suspension in a non-toxic parenterally-acceptable diluent or solvent, for example, as a solution in 1,3-butanediol. Among the acceptable vehicles and solvents which may be employed are water, Ringer's solution, and isotonic sodium chloride solution.

In addition, sterile, fixed oils are conventionally employed as a solvent or suspending medium. For this purpose, any bland fixed oil may be employed, including synthetic mono-or diglycerides. In addition, fatty acids such as oleic acid may be used in the preparation of injectables.

Compounds of formula I may also be administered in the form of liposome delivery systems, such as small unilamellar vesicles, large unilamellar vesicles, and multilamellar vesicles. Liposomes can be formed from a variety of phospholipids, such as cholesterol, stearylamine, or phosphatidylcholines.

Dosage levels of the compound of formula I of the present invention will usually be of the order of about 0.5mg to about 20mg per kilogram body weight, with a preferred dosage range between about 0.5mg to about 10mg per kilogram body weight per day (from about 0.5g to about 3g per patient per day). The amount of active ingredient which may be combined with the carrier materials to produce a single dosage will vary, depending upon the host to be treated and the particular mode of administration. For example, a formulation intended for oral administration to humans may contain about 5mg to 1g of an active compound with an appropriate and convenient amount of carrier material, which may vary from about 5 to 95 percent of the total composition. Dosage unit forms will generally contain between from about 5mg to 500mg of active ingredient.

It will be understood, however, that the specific dose level for any particular patient will depend upon a variety of factors including the activity of the specific compound employed, the age, body weight, general health, sex, diet, time of administration, route of administration, rate of excretion, drug combination and the severity of the particular disease undergoing therapy.

In addition, some of the compounds of the invention may form solvates with water or common organic

solvents. Such solvates are encompassed within the scope of the invention.

The compounds of the invention may additionally be combined with other therapeutic compounds to provide an operative combination. It is intended to include any chemically compatible combination of pharmaceutically-active agents, as long as the combination does not eliminate the activity of the compound of formula I of this invention.

In evaluation of the compounds of the invention, conventional measures of efficacy of treatment of osteoarthritis may be used. For example, commonly-used primary efficacy end-points include the Western Ontario and McMaster's University Osteoarthritis Index (WOMAC) Pain subscale, Patient Global Assessment of Response to Therapy, Investigator Global Assessment of Disease Status, McGill pain questionnaire, Modified Stanford Health Assessment Questionnaire (MHAQ), Health Assessment Questionnaire and Kellgren-Lawrence radiographic grading.

General Methods

Cyclic peptide compounds of formula I are prepared according to methods described in detail in our earlier applications No. PCT/AU98/00490 and PR8334, the entire disclosures of which are incorporated herein by this reference. While the invention is specifically illustrated with reference to the compound AcF-[OPdChaWR] (PMX53), whose corresponding linear peptide is Ac-Phe-Orn-Pro-dCha-Trp-Arg, it will be clearly understood that the invention is not limited to this compound.

Compounds **1-6**, **17**, **20**, **28**, **30**, **31**, **36** and **44** disclosed in International patent application No.PCT/AU98/00490 and compounds **10-12**, **14**, **15**, **25**, **33**, **35**, **40**, **45**, **48**, **52**, **58**, **60**, **66**, and **68-70** disclosed for the first time in Australian provisional application No. PR8334 have appreciable antagonist potency (IC50 < 1 μM)

against the C5a receptor on human neutrophils. PMX53 (compound **17** of PCT/AU98/00490; also identified as compound **1** in PR8334) and compounds **33**, **45** and **60** of PR8334 are most preferred.

We have found that all of the compounds of formula I which have so far been tested have broadly similar pharmacological activities, although the physicochemical properties, potency, and bioavailability of the individual compounds varies somewhat, depending on the specific substituents.

The following general tests may be used for initial screening of candidate inhibitor of G protein-coupled receptors, and especially of C5a receptors.

Receptor-Binding Assay

Assays are performed with fresh human PMNs, isolated as previously described (Sanderson *et al*, 1995), using a buffer of 50 mM HEPES, 1 mM $CaCl_2$, 5 mM $MgCl_2$, 0.5% bovine serum albumin, 0.1% bacitracin and 100 µM phenylmethylsulfonyl fluoride (PMSF). In assays performed at 4°C, buffer, unlabelled human recombinant C5a (Sigma) or peptide, Hunter/Bolton labelled ^{125}I-C5a (~ 20 pM) (New England Nuclear, MA) and PMNs (0.2×10^6) are added sequentially to a Millipore Multiscreen assay plate (HV 0.45) having a final volume of 200 µL/well. After incubation for 60 min at 4°C, the samples are filtered and the plate washed once with buffer. Filters are dried, punched and counted in an LKB gamma counter. Non-specific binding is assessed by the inclusion of 1mM peptide or 100 nM C5a, which typically results in 10-15% total binding.

Data are analysed using non-linear regression and statistics with Dunnett post-test.

Myeloperoxidase Release Assay for Antagonist Activity

Cells are isolated as previously described (Sanderson *et al*, 1995) and incubated with cytochalasin B

(5µg/mL, 15 min, 37°C). Hank's Balanced Salt solution containing 0.15% gelatin and peptide is added on to a 96 well plate (total volume 100 µL/well), followed by 25 µL cells ($4x10^6$/mL). To assess the capacity of each peptide to antagonise C5a, cells are incubated for 5 min at 37°C with each peptide, followed by addition of C5a (100 nM) and further incubation for 5 min. Then 50 µL of sodium phosphate (0.1M, pH 6.8) is added to each well, the plate was cooled to room temperature, and 25 µL of a fresh mixture of equal volumes of dimethoxybenzidine (5.7 mg/mL) and H_2O_2 (0.51%) is added to each well. The reaction is stopped at 10 min by addition of 2% sodium azide. Absorbances are measured at 450 nm in a Bioscan 450 plate reader, corrected for control values (no peptide), and analysed by non-linear regression.

***In Vivo* Assays of Anti-Inflammatory Activity**

The following well-known *in vivo* assay systems are used to assess the anti-inflammatory activity of compounds of the invention. All assay data are analysed using non-linear regression analysis and Student's t-test, analysis of variance, with $p<0.05$ as the threshold level of significance.

(a) Carrageenan Paw Oedema

Anaesthetised (i.p. ketamine & xylazine) Wistar rats (150-200g) or mice are injected with sterilised air (20ml day 1, 10ml day 4) into the subcutaneous tissue of the back. The cavity can be used after 6 days, whereupon carrageenan (2ml, 1% w/w in 0.9% saline) is injected into the air pouch, and exudate is collected after 10 hr. Test compounds are administered daily after Day 6, and their anti-inflammatory effects assayed by differential counting of cells in the air-pouch exudate. Animals are killed at appropriate times after injection, and 2ml 0.9% saline is used to lavage the cavity; lavage fluids are transferred to heparinised tube and cells are counted with a

haemocytometer and Diff-Quik stained cytocentrifuged preparation.

Alternatively, a routine carrageenan paw oedema developed in Wistar rats by administering a pedal injection of carrageenan may be used to elicit oedema which is visible in 2h and maximised in 4h. Test compounds are given 40 min before inflammatory stimulus and evaluated by microcaliper measurements of paws after 2 & 4 hr. See Fairlie,D.P. *et al* (1987). Also see Walker and Whitehouse (1978).

(b) Adjuvant Arthritis.

Adjuvant arthritis is induced in rats (3 strains) either microbially (injection of heat-killed *Mycobacterium tuberculosis*) or chemically (with avridine) by inoculation with the arthritogenic adjuvant co-administered with oily vehicles (Freund's adjuvants) in the tail base. (See Whitehouse, M. W., Handbook of Animal Models for the Rheumatic Diseases, Eds. Greenwald, R. A.; Diamond, H. S.; Vol. 1, pp. 3-16, CRC Press).

Within 13 days the adjuvant arthritis is manifested by local inflammation and ulceration in the tail, gross swelling of all four paws, inflammatory lesions in paws and ears, weight loss and fever. These symptoms, which are similar to those of inflammatory disease in humans (Winter and Nuss, 1966), can be alleviated by agents such as indomethacin or cyclosporin, which also show beneficial effects in man (*eg.* Ward and Cloud, 1966). Without drug treatment at Day 14, arthritic rats had hypertrophy of the paws, reduced albumin but raised acute phase reaction proteins in serum, and depressed hepatic metabolism of xenobiotics as indicated by prolonged barbiturate-induced sleeping times.

To assess activity, compounds are administered for 4 days orally (≤10mg/kg/day) or (intraperitoneally (i.p.) from Days 10-13 following inoculation with arthritogen (Day 0). If the compound is active, the

inflammation is either not visible, or is very significantly reduced in rear or front paws, as assessed by microcaliper measurements of paw thickness and tail volume, as well as by gross inspection of inflammatory lesions. Animals are sacrificed by cervical dislocation on Day 18 unless arthritis signs are absent, whereupon duration of observations is continued with special permission from the Ethics Committees. Experiments are staggered to maximise throughput and allow early comparisons between compounds. This routine assay is well-accepted as identifying anti-inflammatory agents for use in humans.

The invention will now be described by way of reference only to the following general methods and experimental examples.

Example 1 Reverse Passive Arthus Reaction in the Rat

A reverse passive peritoneal Arthus reaction was induced as previously described (Strachan *et al.*, 2000), and a group of rats were pretreated prior to peritoneal deposition of antibody with AcF-[OPdChaWR] (**1**) by oral gavage (10mg kg^{-1} dissolved in 10% ethanol/90% saline solution to a final volume of 200µl) or an appropriate oral vehicle control 30 min prior to deposition of antibody. Female Wistar rats (150-250g) were anaesthetised with ketamine (80mg kg^{-1} i.p.) and xylazine (12mg kg^{-1} i.p.).

The lateral surfaces of the rat were carefully shaved and 5 distinct sites on each lateral surface clearly delineated. A reverse passive Arthus reaction was induced in each dermal site by injecting Evans blue (15 mg kg^{-1} i.v.), chicken ovalbumin (20mg kg^{-1} i.v.) into the femoral vein 10min prior to the injection of antibody. Rabbit anti-chicken ovalbumin (saline only, 100, 200, 300 or 400µg antibody in a final injection volume of 30µL) was injected in duplicate at two separate dermal sites on each

lateral surface of the rat, giving a total of 10 injection sites per rat. Rats were placed on a heating pad, and anaesthetic was maintained over a 4h-treatment period with periodic collection of blood samples. Blood was allowed to spontaneously clot on ice, and serum samples were collected and stored at -20°C. Four hours after induction of the dermal Arthus reaction, the anaesthetised rat was euthanased and a $10mm^2$ area of skin was collected from the site of each Arthus reaction. Skin samples were stored in 10% buffered formalin for at least 10 days before histological analysis using haematoxylin and eosin stain. Additionally, a second set of skin samples were placed in 1mL of formamide overnight, and the absorbance of Evans blue extraction measured at 650nm, as an indicator of serum leakage into the dermis. Figure 1 shows the optical density of dermal punch extracts following intradermal injection of rabbit anti-chicken ovalbumin at 0-400 µg $site^{-1}$ following pretreatment with AcF-[OPdChaWR] intravenously, orally or topically. Data are shown as absorbance at 650nm as a percentage of the plasma absorbance, as mean values ± SEM (*n=3-6*). *indicates a *P value* ≤ 0.05 when compared to Arthus control values.

Rats were pretreated with the C5aR antagonist, AcF-[OPdChaWR] (**1**) as the TFA salt, either intravenously (0.3-1mg kg^{-1} in 200µL saline containing 10% ethanol, 10min prior to initiation of dermal Arthus), orally (0.3-10mg kg^{-1} in 200 µL saline containing 10% ethanol by oral gavage, 30min prior to initiation of dermal Arthus in rats denied food access for the preceding 18hours) or topically (200-400µg $site^{-1}$ 10min prior to initiation of dermal Arthus reaction), or with the appropriate vehicle control. Topical application of the antagonist involved application of 20µl of a 10-20mg mL^{-1} solution in 10% dimethyl sulphoxide (DMSO), which was then smeared directly onto the skin at each site, 10min prior to induction of the Arthus reaction.

The saline-only injection site from rats treated

with Evans blue only served as antigen controls, the saline-only injection site from rats treated with Evans blue plus topical DMSO only served as a vehicle control, the saline-only injection site from rats treated with Evans blue plus either intravenous, oral or topical antagonist only served as antagonist controls, and Evans blue plus dermal rabbit anti-chicken ovalbumin served as antibody controls. Topical application of the peptide AcF-[OPGWR] which has similar chemical composition and solubility as AcF-[OPdChaWR] (**1**), but with an IC_{50} binding affinity of >1mM in isolated human PMNs., served as an inactive peptide control. AcF-[OPGWR] was also dissolved in 10% DMSO and applied topically at 400µg site^{-1} 10 min prior to initiation of the Arthus reaction.

TNFα Measurement

Serum TNFα concentrations were measured using an enzyme-linked immunosorbent assay (ELISA) kit (Strachan *et al.*, 2000). Antibody pairs used were a rabbit anti-rat TNFα antibody coupled with a biotinylated murine anti-rat TNFα antibody. Figure 2 shows the serum TNFα concentrations at regular intervals after initiation of a dermal Arthus reaction, with group of rats pretreated with AcF-[OPdChaWR] intravenously, orally or topically. Data are shown as mean values ± SEM (*n=3-6*). *indicates a *P value* of ≤ 0.05 when compared to Arthus control values.

Interleukin-6 Measurement

An ELISA method as described previously was used to measure serum and peritoneal lavage fluid interleukin-6 (IL-6) concentrations (Strachan *et al.*, 2000).

Pathology Assessment

Rat skin samples were fixed in 10% buffered formalin for at least 10 days, and stained with haematoxylin and eosin using standard histological techniques. Dermal samples were analysed in a blind

fashion for evidence of pathology, and the degree of rat PMN infiltration was scored on a scale of 0-4. Initiation of a dermal Arthus reaction resulted in an increase in interstitial neutrophils, which was quantified in the following manner. Sections were given a score of 0 if no abnormalities were detected. A score of 1 indicated the appearance of increased PMNs in blood vessels, but no migration of inflammatory cells out of the lumen. A score of 2 and 3 indicated the appearance of increasing numbers of PMNs in the interstitial tissue and more prominent accumulations of inflammatory cells around blood vessels. A maximal score of 4 indicated severe pathological abnormalities were present in dermal sections, with excessive infiltration of PMNs into the tissues and migration of these cells away from blood vessels. Figure 3 shows the intradermal injection of increasing amounts of antibody leads to a dose-responsive increase in the pathology index scored by dermal samples (A). Data are shown for dermal samples intradermally injected with saline or 400 µg site^{-1} antibody (n=5) in rats pretreated with AcF-[OPdChaWR] intravenously (B) (n=3), orally (C) (n=3) and topically (D) (*n=3*). Data are shown as mean values ± SEM. * *P≤0.05* when compared to Arthus values using a non-parametric t-test.

Example 2 Rat Monoarticular Antigen-Induced Arthritis

Female Wistar rats (150-250g) were obtained from the Central Animal Breeding House, University of Queensland. Methylated bovine serum albumin (mBSA) (0.5mg) was dissolved in Freund's complete adjuvant (0.5mg) and sonicated to produce a homogenous suspension. Each rat received a subcutaneous injection of this suspension (0.5mL) on days 1 and 7. On day 12-28, rats were separated into separate cages, and body weight and food and water intake monitored daily. Rats received

either ordinary tap water or drinking water containing AcF-[OPdChaWR] (1). Body weight and water intake were monitored daily, and rats received a daily dose of 1mg/kg/day of the C5aR antagonist AcF-[OPdChaWR] (1) for days 12-28 of the trial. On day 14, rats were anaesthetised and their hind limbs shaven. Each rat received an intra-articular (100µl) injection of mBSA (0.5mg) in the left knee, and saline in the right knee. The saline only knee from rats receiving normal drinking water served as a saline control, the saline only knee from rats receiving AcF-[OPdChaWR] (1) in the drinking water served as an antagonist control.

Rats were euthanased on day 28, and whole blood collected into an Eppendorf tube and allowed to spontaneously clot on ice. Blood samples were centrifuges (11,000 rpm x 3min) and serum collected and stored at -20°C until analysis of serum cytokines using an ELISA. Each knee capsule was lavaged with 100µL saline, and the total cell count determined using a haemocytometer. In addition, an aliquot of the knee joint lavage fluid was dropped onto a glass slide, and allowed to air dry. Once dry, cells were stained with a differential stain (Diff Quick) and a differential cell count was performed using a 40 x dry lens microscope. The remaining lavage fluids from each joint were stored at -20°C until later analysis of intra-articular cytokine levels using an ELISA. Each knee joint was severed, trimmed leaving only the area of interest and the skin was split with a scalpel blade. Knee samples were stored in 10 % buffered formalin for ≥10 d. Knees were then rinsed with distilled water and placed in a saturated solution of EDTA solution for 21 d for decalcification before being embedded in paraffin wax.

Knee tissue samples were prepared using standard histological techniques as described above in Example 6 and stained using an heamotoxylin and eosin stain. Histological slides were analysed in a blind fashion. Tissue sections were scored from 0-4, with a score of 0 indicating the detection of no abnormalities, and increasing scores with the appearance of synovial cell proliferation, inflammatory cell infiltration, cartilage destruction and haemorrhage. In no samples was there evidence of significant bone erosion. Samples were thawed on the day of ELISA analysis, and serum or intra-articular lavage fluid TNFα and IL-6 concentrations were determined from a standard curve, using an ELISA as described previously in Example 6.

Figure 4 shows the inhibition of arthritic right knee joint swelling by AcF-[OPdChaWR] given orally on Days-2 to +14, while Figure 5 shows the inhibition of right knee joint TNF-α and IL-6 levels in joint lavage. Untreated refers to animal not treated with AcF-[OPdChaWR] but with right knee challenged with antigen following sensitisation.

Example 3 The effect of PMX53 on induced osteoarthritis in dogs

Four healthy dogs were obtained from a group of pound dogs destined for euthanasia. The dogs were acclimatised to the experimental canine ward for 7 days before the experiment commenced. Dogs were housed individually and fed on a mixture of dry dog food, canned dog food and occasional raw bones. They were walked for 10 minutes twice daily throughout the trial. The dogs were of age and sex as follows:

Dog #2 Male cattle dog (approximately 1 year old)

Dog #3 Female cattle dog (1 - 2 years old)
Dog #4 Female bull terrier cattle dog cross (2 years old)
Dog #5 Female whippet cross (less than 1 year old)

PMX53 was synthesised by the Institute of Molecular Bioscience at the University of Queensland. The drug was dissolved in 30% polyethylene glycol 400 (PEG 400) in normal saline. The solution was made to a concentration of 3mg/ml. The placebo vehicle was 30% PEG 400 in saline. All treatments were sterilised by filtration and stored at 4°C. The dogs were given the drug or placebo at a dose rate of 1ml/10kg body weight. The treatments were assigned to the dogs in a random fashion. The drug containers were so marked that the attendants who scored the gait were unaware of the treatment given to each dog.

The cruciate ligament of the left stifle (knee) joint was transected surgically by an experienced specialist surgeon. The incision in the skin was approximately 2 centimetres long. The dogs were given postoperative pain relief, including epidural analgesia and opioids. At 24 hours after the surgery all dogs were limping. No supplementary analgesic therapy was deemed necessary after day 1. All dogs tolerated the procedure very well.

The drug was given as a subcutaneous injection (0.3 mg/kg) into the loose skin on the dorsum of the neck once daily. The drug and placebo caused mild discomfort when injected, but the dogs were readily distracted with food at the time of injection. Dogs were assessed twice daily for lameness while they were walked. The scoring was done by at least two people.

The dogs treated with placebo showed an improvement in gait between 10 and 13 days after surgery, as shown in Figure 6a. This was seen as a natural recovery from the injury. They never returned to complete

soundness.

In contrast to the controls, drug-treated dogs showed improvement in the degree of lameness after 4 - 6 days' treatment, ie more rapidly than controls, as illustrated in Figure 6b. The dogs maintained the improved gait for the next 3 weeks until the end of the trial. These two dogs were relatively sound at the end of the trial.

The dogs were euthanased at day 28. All dogs had complete disruption of the anterior cruciate ligament in the left knee, and all had thickening of the joint capsule and increased volume of joint fluid in the left knees. There were tags of fibrin adhering to the synovium in all affected joints. There was mild cartilage erosion in dog #2 (drug treated) and dog #4 (placebo). The site of injection showed no abnormalities.

The joints were X-rayed before surgery and after death. No significant bony lesions were detected at either time point. Blood was collected for biochemistry and haematology before surgery and at day 28. No abnormalities were detected in any dog at either time point. The histopathology of the joint showed that there was reduced thickening of the synovial membrane and reduced fibrosis of the drug-treated dogs compared to the arthritic controls. This is illustrated in Figure 7. The clinical impression was the drug clearly improved the ability of the dogs to walk. The drug-treated dogs walked strongly on the affected leg from approximately one week after surgery and the commencement of treatment. It should be noted that both these dogs had a lower starting score than the two placebo-treated dogs.

The placebo-treated dogs improved slowly, so by 4 weeks they were noticeably less lame than at the start of the trial. This is consistent with practice observations; small animals with ruptured cruciate ligaments frequently recover without surgery.

DISCUSSION

Cyclic peptides have several important advantages over acyclic peptides as drug candidates (Fairlie et al 1995, Fairlie et al, 1998, Tyndall and Fairlie, 2001). The cyclic compounds described in this specification are stable to proteolytic degradation for at least several hours at 37°C in human blood or plasma, in human or rat gastric juices, or in the presence of digestive enzymes such as pepsin, trypsin and chymotrypsin. In contrast, short linear peptides composed of L-amino acids are rapidly degraded to their component amino acids within a few minutes under these conditions. A second advantage lies in the constrained single conformations adopted by the cyclic and non-peptidic molecules, in contrast to acyclic or linear peptides, which are flexible enough to adopt multiple structures in solution other than the one required for receptor-binding. Thirdly, cyclic compounds such as those described in this invention are usually more lipid-soluble and more pharmacologically bioavailable as drugs than acyclic peptides, which can rarely be administered orally. Fourthly, the plasma half-lives of cyclic molecules are usually longer than those of peptides.

It will be apparent to the person skilled in the art that while the invention has been described in some detail for the purposes of clarity and understanding, various modifications and alterations to the embodiments and methods described herein may be made without departing from the scope of the inventive concept disclosed in this specification.

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Figure 1

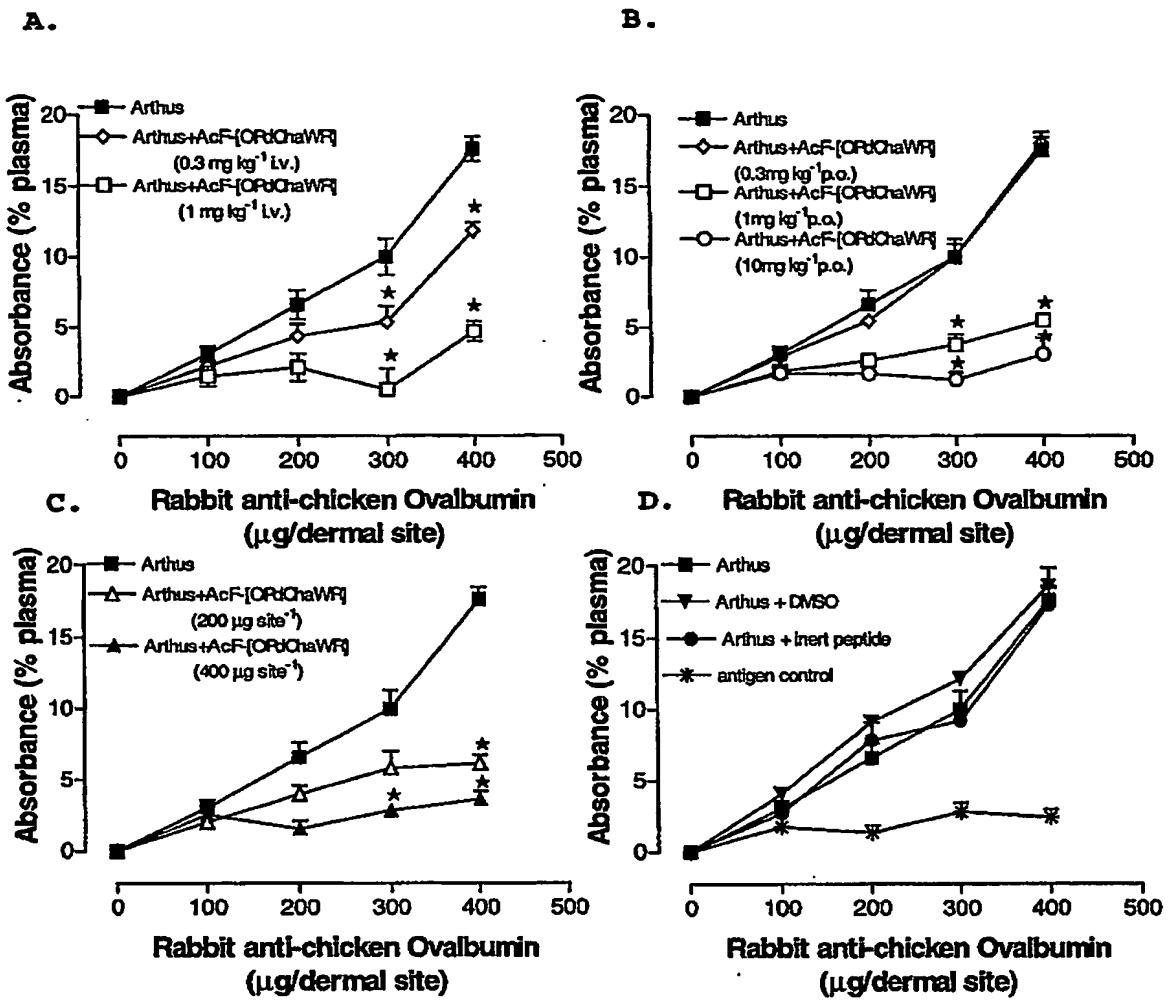

A.
B.
Absorbance (% plasma)
Arthus
Arthus+AcF-[ORdChaWR] (0.3 mg kg-1 i.v.)
Arthus+AcF-[ORdChaWR] (1 mg kg-1 i.v.)
Arthus+AcF-[ORdChaWR] (0.3mg kg-1p.o.)
Arthus+AcF-[ORdChaWR] (1mg kg-1p.o.)
Arthus+AcF-[ORdChaWR] (10mg kg-1p.o.)
Rabbit anti-chicken Ovalbumin (µg/dermal site)
C.
D.
Arthus+AcF-[ORdChaWR] (200 µg site-1)
Arthus+AcF-[ORdChaWR] (400 µg site-1)
Arthus + DMSO
Arthus + inert peptide
antigen control

Figure 2

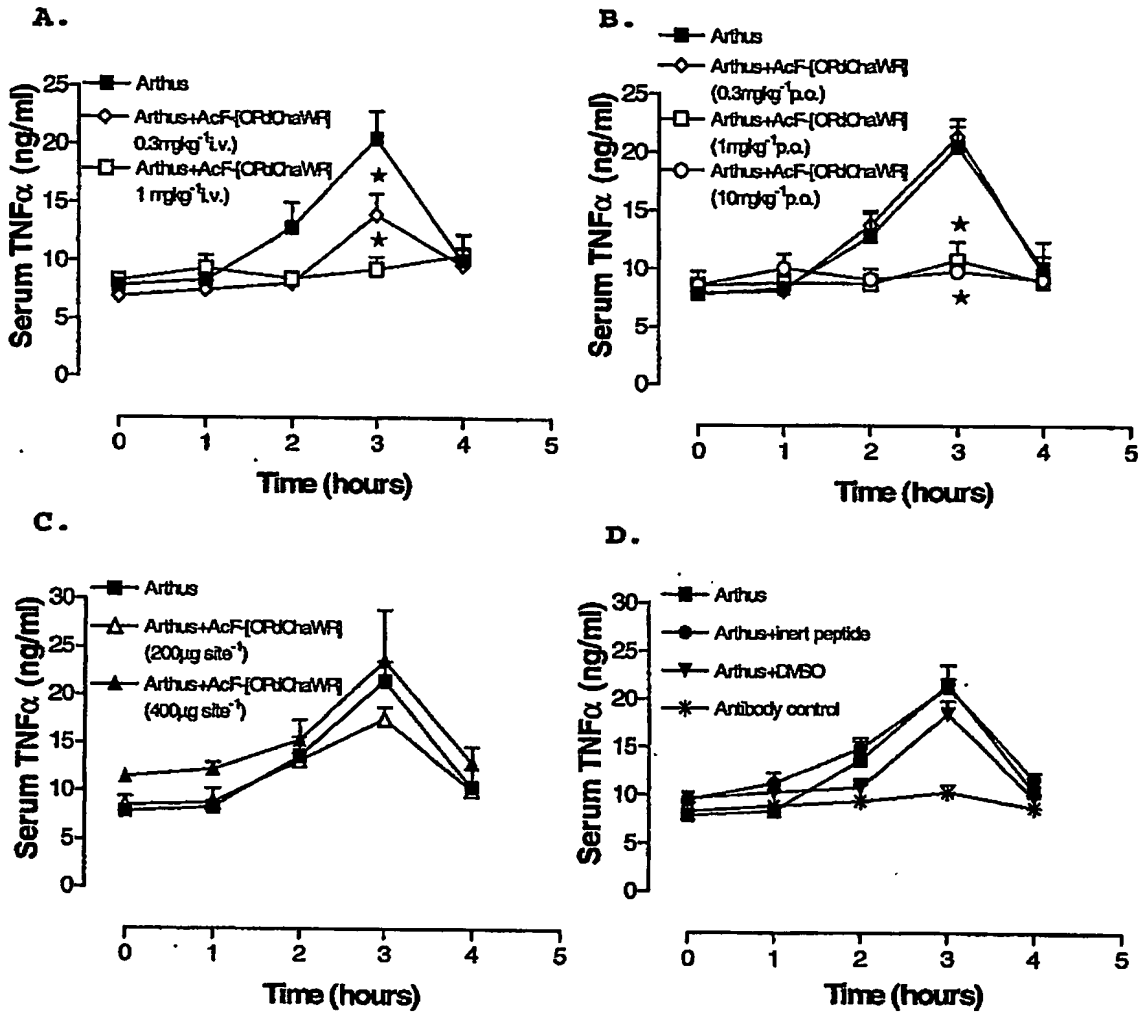

A.
Arthus
Arthus+AcF-[ORdChaWR] 0.3mg/kg⁻¹ i.v.)
Arthus+AcF-[ORdChaWR] 1 mg/kg⁻¹ i.v.)
Serum TNFα (ng/ml)
Time (hours)
B.
Arthus
Arthus+AcF-[ORdChaWR] (0.3mg/kg⁻¹ p.o.)
Arthus+AcF-[ORdChaWR] (1mg/kg⁻¹ p.o.)
Arthus+AcF-[ORdChaWR] (10mg/kg⁻¹ p.o.)
Serum TNFα (ng/ml)
Time (hours)
C.
Arthus
Arthus+AcF-[ORdChaWR] (200µg site⁻¹)
Arthus+AcF-[ORdChaWR] (400µg site⁻¹)
Serum TNFα (ng/ml)
Time (hours)
D.
Arthus
Arthus+inert peptide
Arthus+DMSO
Antibody control
Serum TNFα (ng/ml)
Time (hours)

Figure 3

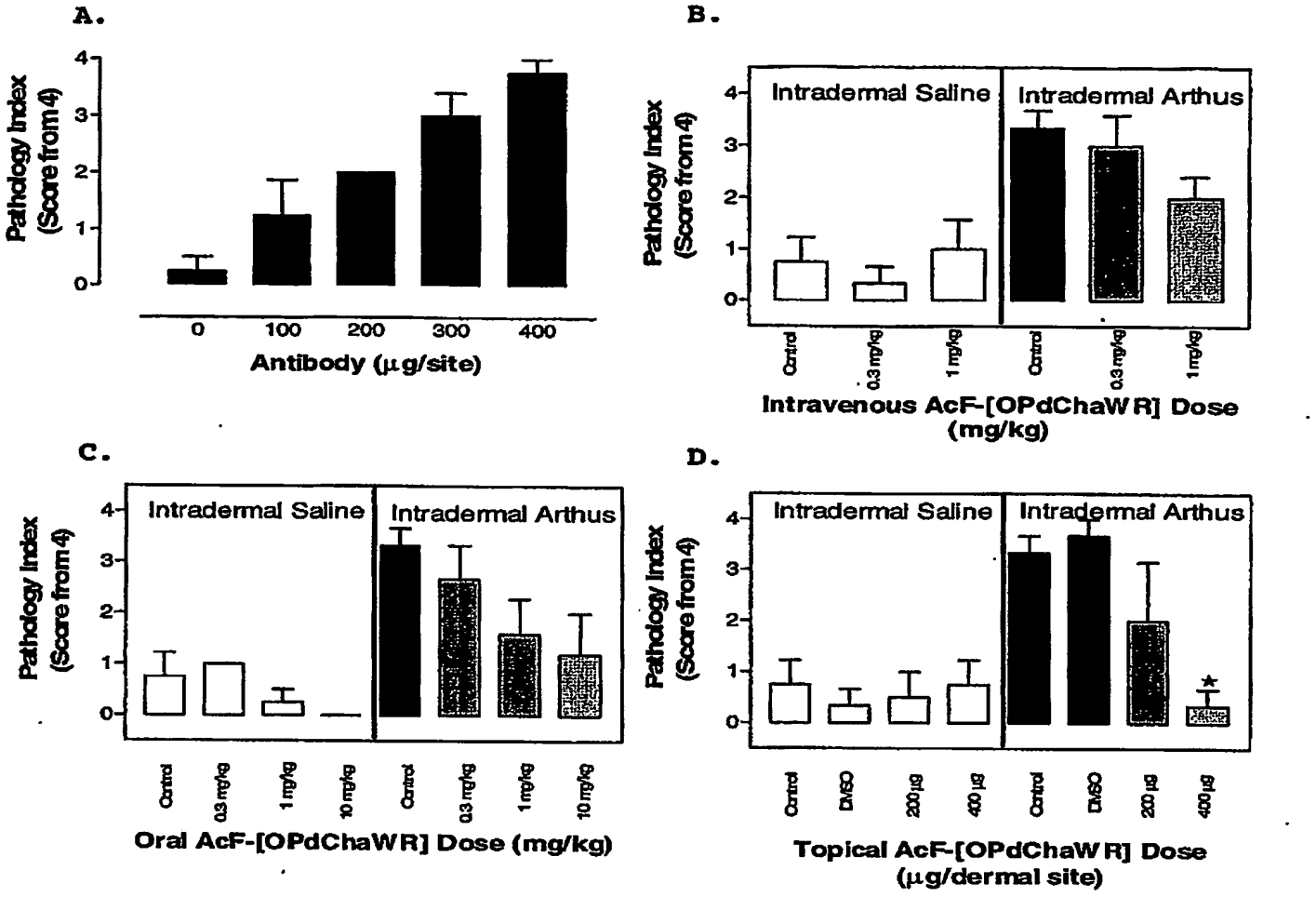

A.
Pathology Index (Score from 4)
0
1
2
3
4
0
100
200
300
400
Antibody (µg/site)
B.
Intradermal Saline
Intradermal Arthus
Pathology Index (Score from 4)
Control
0.3 mg/kg
1 mg/kg
Control
0.3 mg/kg
1 mg/kg
Intravenous AcF-[OPdChaWR] Dose (mg/kg)
C.
Intradermal Saline
Intradermal Arthus
Pathology Index (Score from 4)
Control
0.3 mg/kg
1 mg/kg
10 mg/kg
Control
0.3 mg/kg
1 mg/kg
10 mg/kg
Oral AcF-[OPdChaWR] Dose (mg/kg)
D.
Intradermal Saline
Intradermal Arthus
Pathology Index (Score from 4)
Control
DMSO
200 µg
400 µg
Control
DMSO
200 µg
400 µg
Topical AcF-[OPdChaWR] Dose (µg/dermal site)

Figure 4

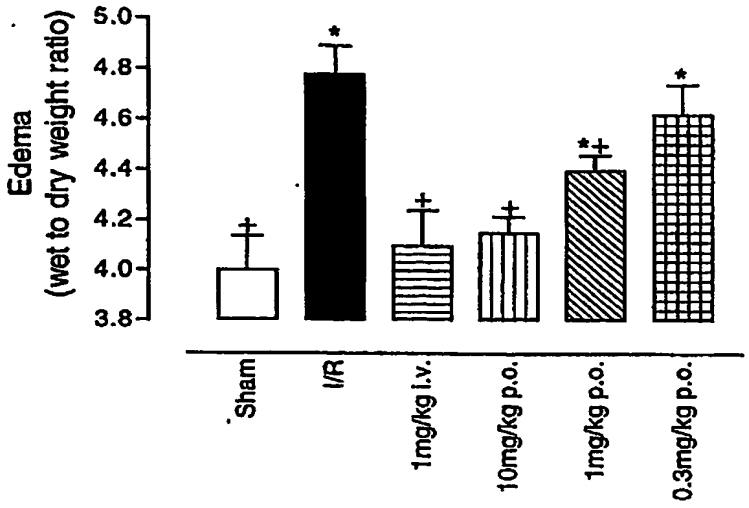


Figure 5

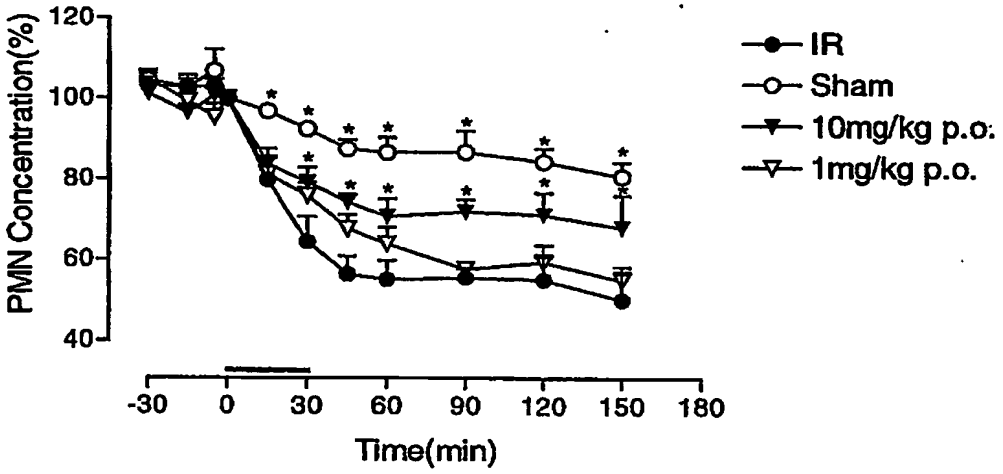


Figure 6a

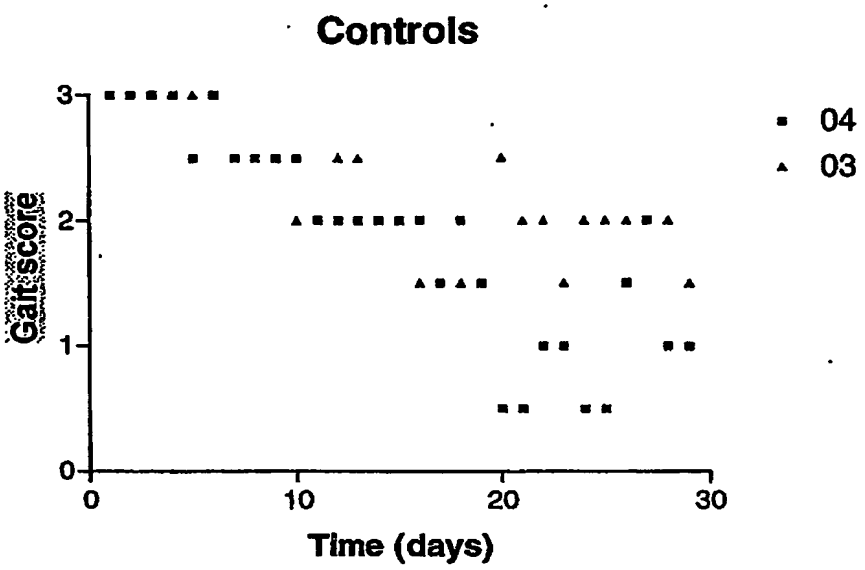


Figure 6b

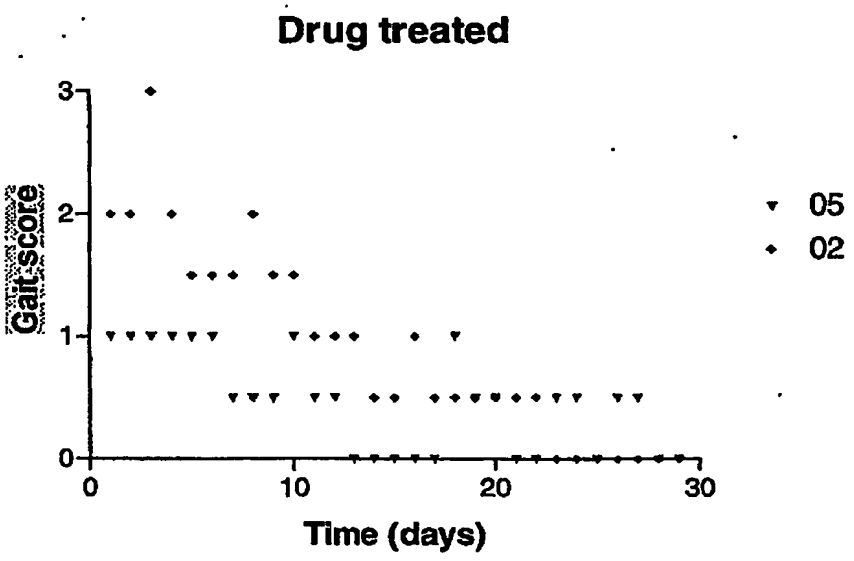
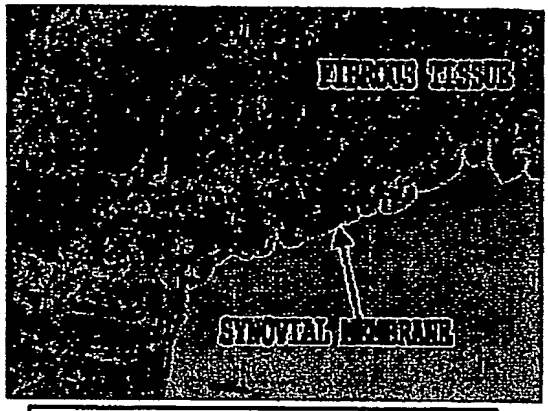
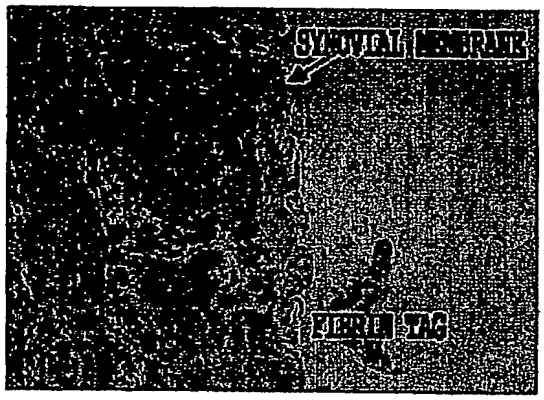


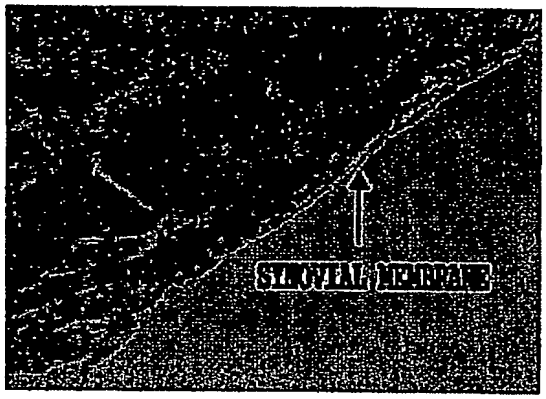
Figure 7

CONTROL DOG KNEE JOINT



CONTROL DOG KNEE JOINT





3D53-TREATED (0.3 MG/KG/DAY S.C.) KNEE JOINTS. DRUG TREATED SPECIMENS SHOW REDUCED THICKENING OF SYNOVIAL MEMBRANE WITH LESS INFLAMMATORY CELL INFILTRATE, AS WELL AS REDUCED ACCUMULATION OF FIBROUS TISSUE